

# Radiation dosimetry through statistical analysis of biomarkers

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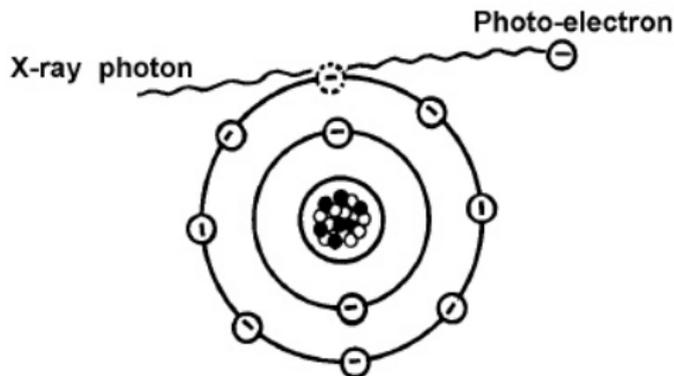


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- ▶ When we talk about radiation, we often mean **ionizing radiation** ( $\alpha$  and  $\beta$  particles,  $\gamma$ -rays, X-rays, neutrons...), which carries enough energy to ionize atoms or molecules. Ionizing radiation can cause serious damage to cells, tissues, and DNA.



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- ▶ A **biomarker** (short for: “biological marker”) is “any substance, structure, or process that can be measured in the body or its products and influence or predict the incidence of outcome or disease” (WHO).
- ▶ **Biodosimetry** is a dosimetry technique which exploits the information provided by radiation-sensitive biomarkers (usually, radiation-induced damage inside the cell nucleus) to infer the radiation dose.

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- ▶ **Biodosimetry** is a dosimetry technique which exploits the information provided by radiation-sensitive biomarkers (usually, radiation-induced damage inside the cell nucleus) to infer the radiation dose.
- ▶ **Statistics** comes into play when establishing the link between the biomarker and the absorbed dose.

# Radiation biodosimetry

- ▶ Radiation accident or incident leading to irradiated blood lymphocytes.
- ▶ Need rapid and reliable procedures to determine the radiation dose contracted by individuals.
- ▶ Members of the public do not usually wear radiation dosimeters...
- ▶ Hence, triage and clinical decision making needs to rely on biomarkers to estimate the contracted radiation dose.



## Cytogenetic biomarkers

- ▶ Example: Frequencies of dicentrics (= aberrant chromosome having two centromeres) in  $n = 4400$  lymphocytes after *in vitro* 'whole body' exposure with 200 kV X-rays.

$x_i$	$y_{ij}$								$n_i$
	0	1	2	3	4	5	6	7	
1	1715	268	15	2	0	0	0	0	2000
2	638	298	56	8	0	0	0	0	1000
3	247	225	85	37	6	0	0	0	600
4	99	129	92	52	21	5	2	0	400
5	48	88	97	99	36	25	5	2	400



- ▶  $x_i$ : dose (in Gy) used to irradiate blood sample  $i$ ,  $i = 1, \dots, 5$ .
- ▶  $y_{ij}$ : counts of dicentric aberrations in  $j$ -th cell of blood sample  $i$ ,  $j = 1, \dots, n_i$ .

## Dose-response modelling

- ▶ These are count data, so a natural choice for the response distribution is Poisson, that is

$$f(y_{ij}|x_i) = e^{-\lambda_i} \frac{\lambda_i^{y_{ij}}}{y_{ij}!}$$

- ▶ There is agreement in the biodosimetry literature that the Poisson means  $\lambda_i$  can be described by a parametric model

$$\lambda_i = E(y_{ij}|x_i) = \beta_0 + \beta_1 x_i + \beta_2 x_i^2$$

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- ▶ For parameter estimation, firstly set up the likelihood function:

$$L = \prod_{i,j} f(y_{ij}|x_i) = \prod_{i,j} e^{-\lambda_i} \frac{\lambda_i^{y_{ij}}}{y_{ij}!} \propto \prod_i e^{-n_i \lambda_i} \lambda_i^{\sum_j y_{ij}}$$

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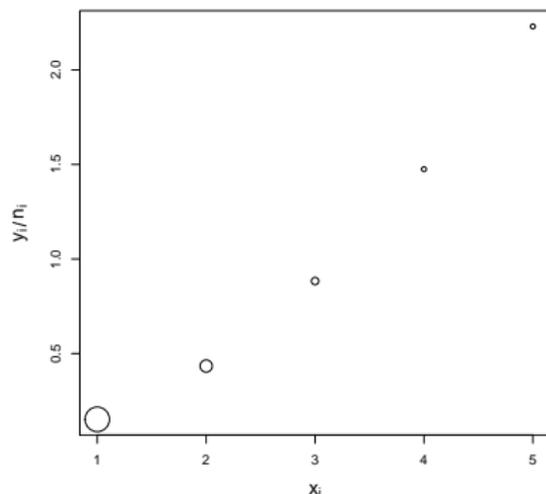
- ▶ One can conveniently work at the *aggregated data* level, with data  $(x_i, y_i) = (x_i, \sum_{j=1}^{n_i} y_{ij})$ .

## Aggregated data

- ▶ Let  $y_i = \sum_j y_{ij}$ . Then the aggregated data are

$x_i$	$n_i$	$y_i$
1.0	2000	304
2.0	1000	434
3.0	600	530
4.0	400	590
5.0	400	892

- ▶ Graphically, with circle size  $\propto n_i$ .



- ▶ 'Empirical dose-response curve'

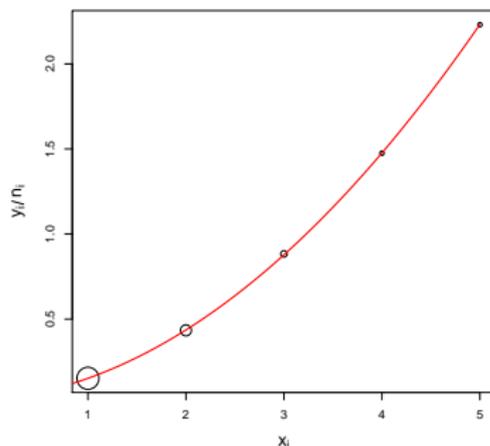
# Poisson regression

- ▶ Model in terms of aggregated data:

$$\lambda_i \equiv E(y_i|x_i)/n_i = \beta_0 + \beta_1 x_i + \beta_2 x_i^2$$

where  $y_i \sim \text{Pois}(n_i \lambda_i)$ .

- ▶ Poisson regression model fitted via ML
  - ▶ special case of generalized linear models
- ▶ Fitted **dose-response curve**,  $\hat{\lambda}_i = \hat{\beta}_0 + \hat{\beta}_1 x_i + \hat{\beta}_2 x_i^2$ :



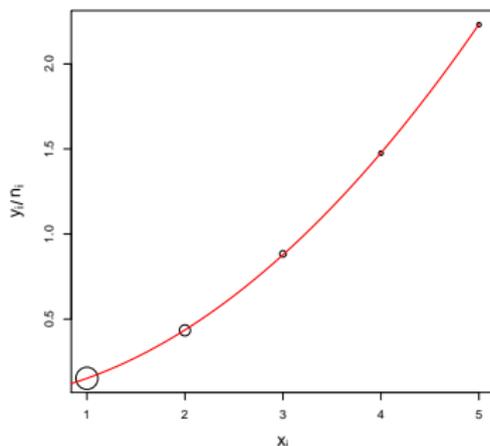
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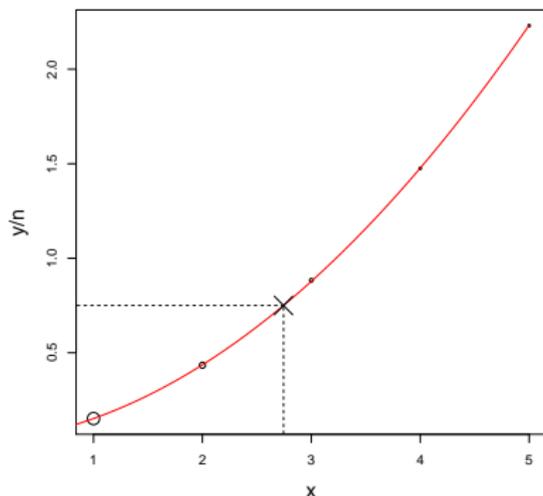
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- ▶ Fitted **dose-response curve**,  $\hat{\lambda}_i = \hat{\beta}_0 + \hat{\beta}_1 x_i + \hat{\beta}_2 x_i^2$ :



- ▶ serves as calibration curve for the dose-estimation problem.

# Inverse regression

- ▶ Dose estimation is an inverse regression problem:
  - ▶ We have a model for the dicentric count,  $y_i$ , given dose  $x_i$ .
  - ▶ In practice, we want to estimate  $x_i$  given  $y_i$ !
- ▶ For instance, assume a patient has been admitted to hospital due to potential radiation exposure. A sample of  $n_0 = 200$  lymphocytes was analyzed, yielding  $y_0 = 150$  dicentrics.



## Dose estimation from calibration curve

- ▶ Mathematically, this is not a big problem. Assume the observed ratio of dicentrics is  $R = \frac{y_0}{n_0}$ . Then we have

$$R = \hat{\beta}_0 + \hat{\beta}_1 x + \hat{\beta}_2 x^2$$

which can be solved wrt  $x$  as

$$\hat{x} = \frac{-\hat{\beta}_1 + \sqrt{\hat{\beta}_1^2 - 4\hat{\beta}_2(\hat{\beta}_0 - R)}}{2\hat{\beta}_2}$$

- ▶ With  $R = \frac{150}{200} = 0.75$ , this gives

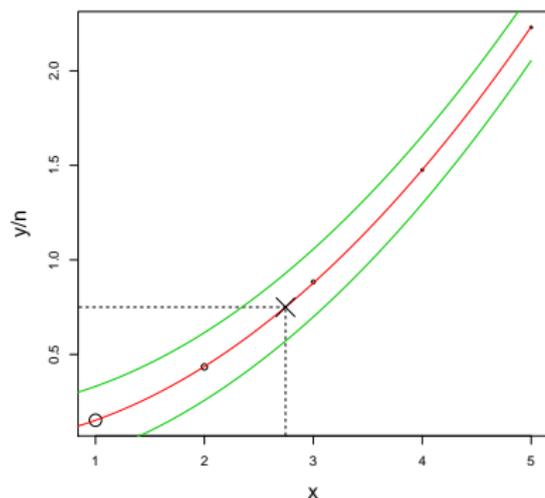
$$\hat{x} = 2.745.$$

# Uncertainty

- ▶ Of course, this estimation is not *exact*.
- ▶ Can one specify the uncertainty in this process?

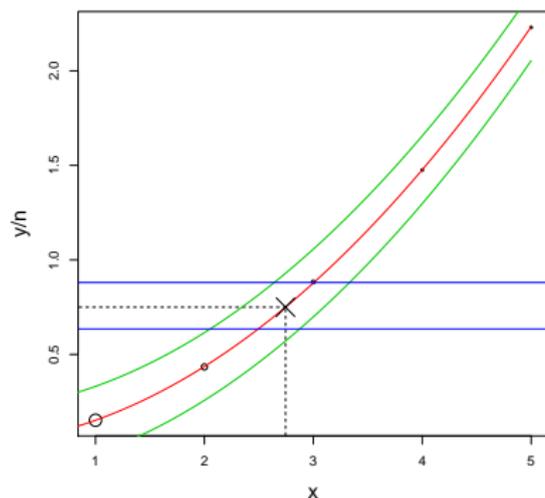
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- ▶ Two sources of uncertainty:
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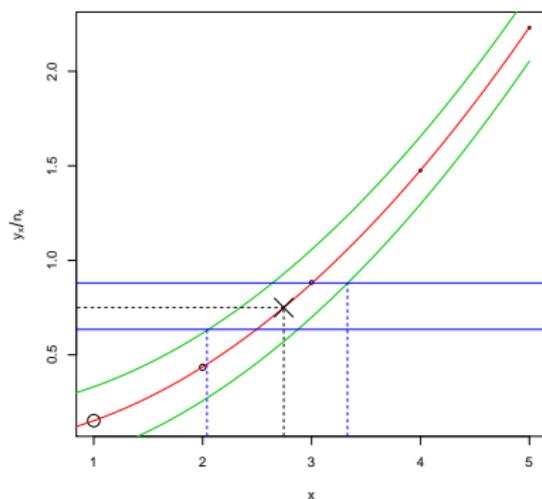
# Uncertainty

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- ▶ Two sources of uncertainty:
  - ▶ Uncertainty due to the **estimation** of the curve (randomness of the calibration data).
  - ▶ Uncertainty due to **random variation** of  $y_0$ , given  $x_0$ .



# Uncertainty bounds

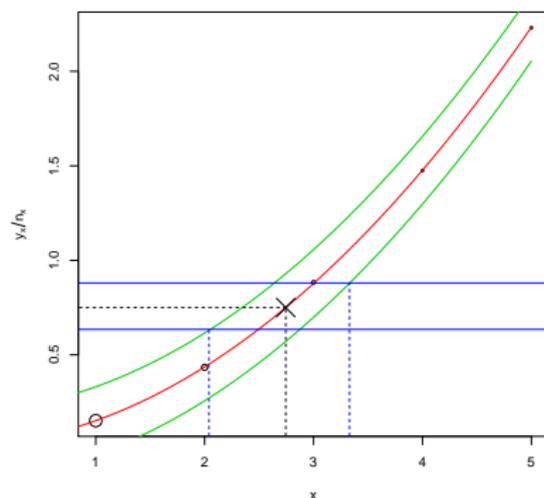
- ▶ Combine the two sources of uncertainty ('Merkle's method', 1983):



- ▶ Here, a 95% confidence interval for the 'true' dose,  $x_0$ , is given as  $[2.04, 3.33]$ .

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- ▶ Here, a 95% confidence interval for the 'true' dose,  $x_0$ , is given as  $[2.04, 3.33]$ .
- ▶ Official uncertainty assessment routine suggested by the International Atomic Energy Agency [IAEA].

## A (semi-)Bayesian approach to uncertainty assessment

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$$L(y_0|\lambda, n_0) \propto e^{-n_0\lambda} \lambda^{y_0}$$

where  $\lambda = \beta_0 + \beta_1x + \beta_2x^2$ , and  $x$  the (true, unknown) dose.

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- ▶ Integration over  $\lambda$  gives *calibrative density of x*:

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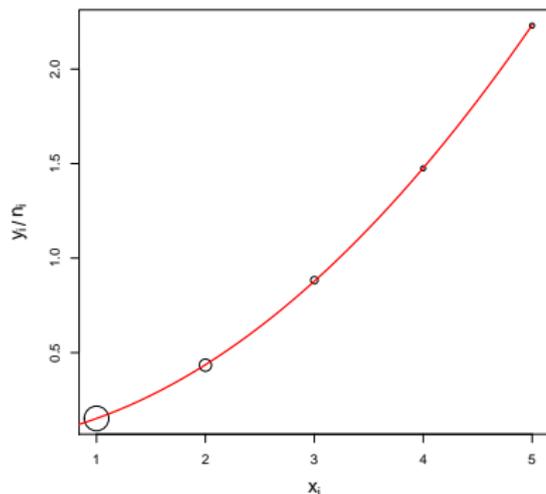
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$$p(x|y_0) \propto p(x) \int L(y_0|\lambda, n_0)\phi(\lambda|x) d\lambda$$

- ▶ Integral has explicit solution via Hermite distribution (Higuera et al, 2015)

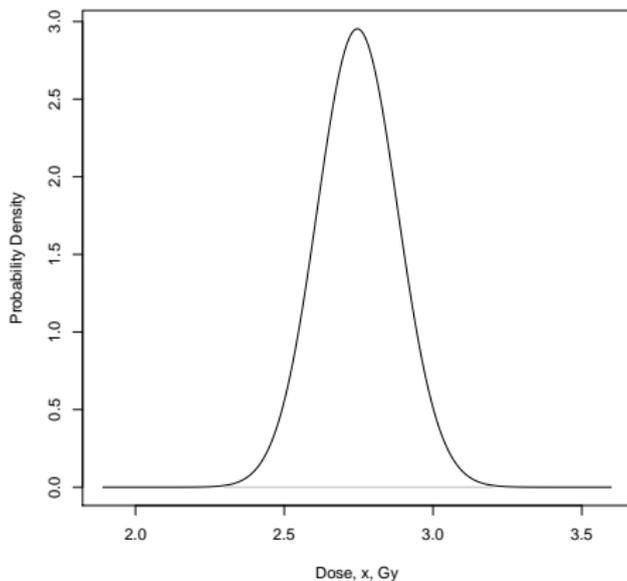
## A (semi-)Bayesian approach to uncertainty assessment

- ▶ Consider again the example before: Patient sample with  $n_0 = 200$ ,  $y_0 = 150$ .
- ▶ Use **the same** estimated dose-response curve,  $\hat{\lambda}_i = \hat{\beta}_0 + \hat{\beta}_1 x_i + \hat{\beta}_2 x_i^2$ , as before:



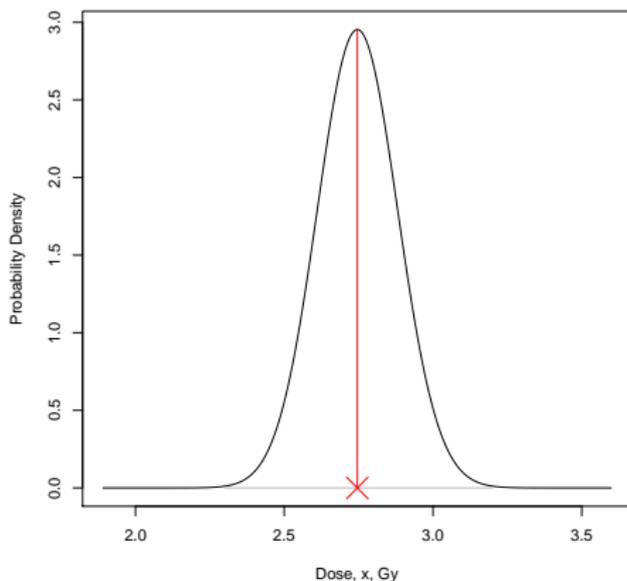
# A (semi-)Bayesian approach to uncertainty assessment

- ▶ Calibrative density for 'true' dose  $x$ , using R package **radir**:



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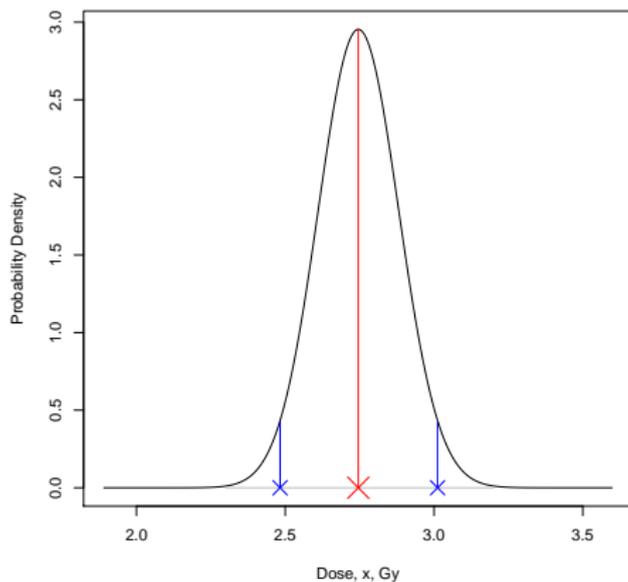
- ▶ Dose estimate: Mode of calibrative density:



- ▶  $\hat{x} = 2.75$ .

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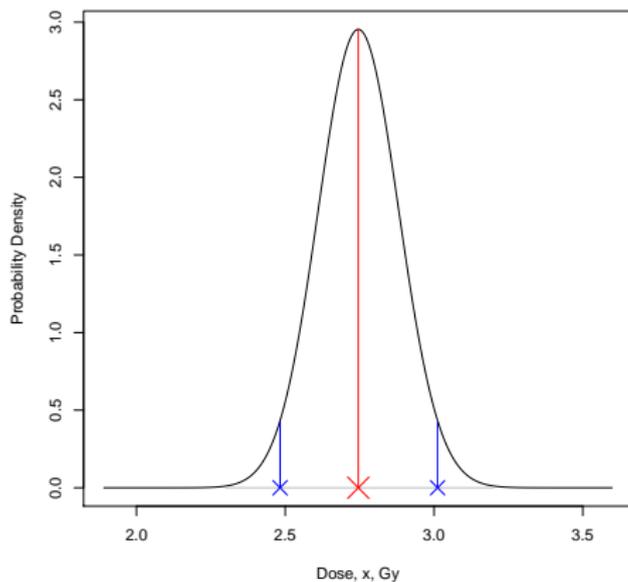
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- $CI = [2.48, 3.01]$

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- $CI = [2.48, 3.01] \in [2.04, 3.33]$

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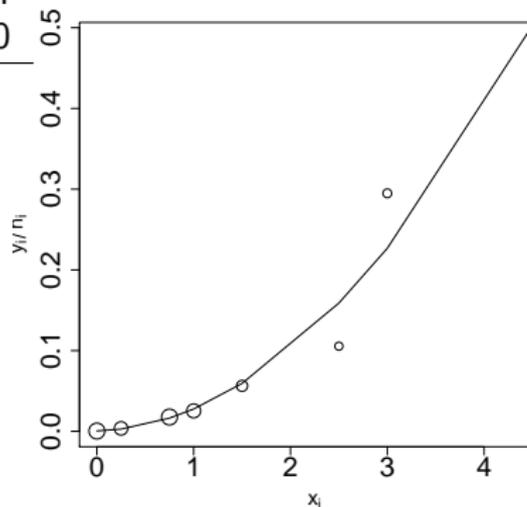
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- ▶ NIHR-funded project (2014): Identification of appropriate response models for chromosomal aberration counts.
- ▶ Initially, carried out an extensive analysis of 11 *in vitro* calibration data sets.
- ▶ It turned out that the data set shown initially in this talk was **the only one** for which the Poisson assumption is (approximately) adequate.
- ▶ In most occasions, things are not quite as nice...
  - ▶ There is **overdispersion** (variance  $\gg$  mean)
  - ▶ There is **zero-inflation** (more zero counts than one would expect under the Poisson model)

## Example: Cobalt-60 $\gamma$ - rays

$x_i$	$y_{ij}$						$n_i$	$y_i$
	0	1	2	3	4	5		
0.00	2591	1	0	0	0	0	2592	1
0.25	2185	8	0	0	0	0	2193	8
0.75	2550	44	1	0	0	0	2595	46
1.00	2231	54	2	0	0	0	2287	58
1.50	1712	96	3	0	0	0	1811	102
2.50	1196	123	7	1	0	0	1327	140
3.00	1070	320	41	6	1	0	1438	424
4.50	895	360	110	25	5	1	1396	680



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- ▶ ...

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- ▶ Negative Binomial model (NB)
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  - ▶ ...
- ▶ However, the presence/type of violation of the Poisson model will often not be obvious (from the fitted calibration curve).
- ▶ Tools which can be used to assist in this question:
  - ▶ Model selection criteria, e.g AIC, BIC
  - ▶ Statistical tests, e.g. LR tests, Score tests
  - ▶ Diagnostic plot

## Model selection

- ▶ Assume we we need to decide between one of the response distributions Poisson, ZIP, NB, ZINB.
- ▶ Use Akaike Information Criterion: Find model which minimizes  $AIC = -2 \log L + 2p$  (Goodness-of-fit/ complexity trade-off).

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- ▶ For instance, for the two data sets displayed so far,

AIC for...	X-rays	$\gamma$ -rays
Poisson	<b>7622.55</b>	7504.73
Zero-inflated Poisson (ZIP)	7623.57	7490.36
Negative Binomial (NB)	7624.56	<b>7489.10</b>
ZINB	7626.39	7491.44

- ▶ For the X-rays, the Poisson model well supported.
- ▶ For the  $\gamma$ -rays, evidence for NB or perhaps ZIP model.

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- ▶ For the X-rays, the Poisson model well supported.
  - ▶ For the  $\gamma$ -rays, evidence for NB or perhaps ZIP model.
- ▶ Model selection methods...
  - ▶ give a useful *indication* of a suitable model;
  - ▶ are easily implemented;
  - ▶ do *not* tell whether one model is *significantly* better than another.

## Score test

- ▶ Say  $H_0 = Po(\lambda_i)$ ,  $H_1 = ZIP(p, \lambda_i)$ ,
  - ▶ in other words,  $H_0 : p = 0$ .
- ▶ Score test statistic:

$$T = S(0, \hat{\beta})^T J(0, \hat{\beta})^{-1} S(0, \hat{\beta})$$

where  $S = \partial L / \partial \beta$  and  $J = \partial L / \partial \beta \beta^T$  are the score function and Fisher information of model  $H_1$ , evaluated under the model fit  $\beta$  under  $H_0$ .

- ▶ Critical value for  $\alpha = 0.05$  is  $\chi_{1,0.95}^2 = 3.84$ .

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- ▶ Critical value for  $\alpha = 0.05$  is  $\chi_{1,0.95}^2 = 3.84$ .
- ▶ Results for 8 data sets (Oliveira et al, 2016):

LET	Homogenous exposure				Partial exposure			
	low	high	low	high	low	high	low	high
id	0.92	18.17	87.72	61.32	2007.39	1418.28	416.20	387.91

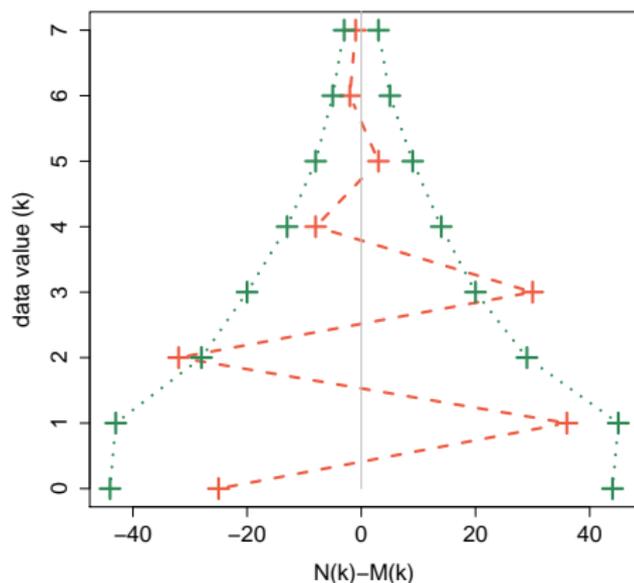
- ▶ (the first two ones are the data sets discussed previously)
- ▶ All data sets except the first one (X-rays) are zero-inflated!!

## New graphical device

- ▶ We developed a new graphical tool to detect zero-inflation (and in fact, any-number-inflation).
- ▶ Effectively, it is demonstrated whether the number of counts, for  $k = 0, 1, 2, \dots$ , is consistent with the specified count distribution.

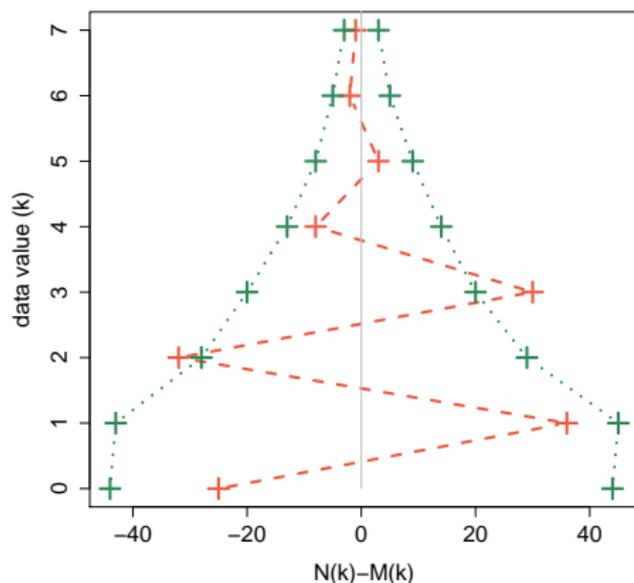
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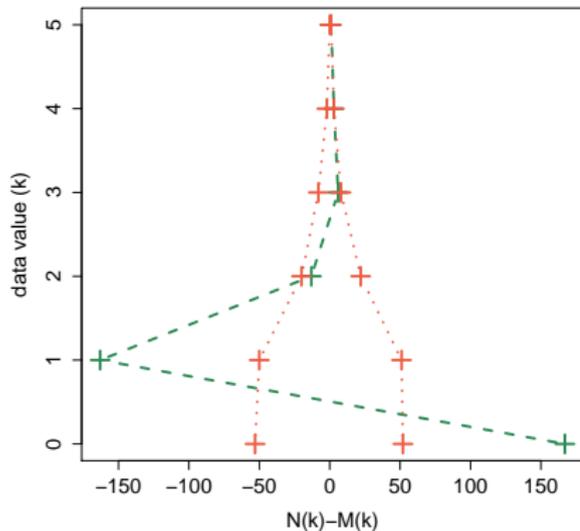
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'Christmas tree plot'  
(Einbeck & Wilson, 2016)

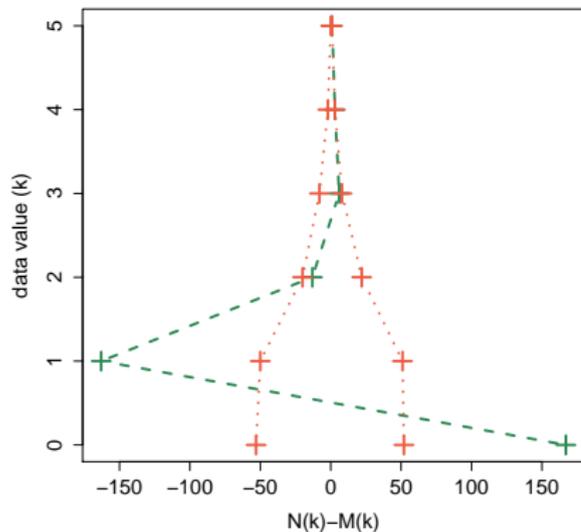
# Christmas tree plot for $\gamma$ -ray data

- ▶ using Poisson model

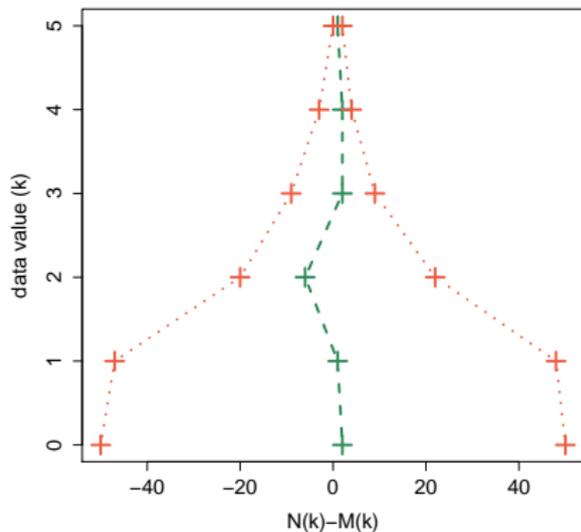


# Christmas tree plot for $\gamma$ -ray data

▶ using Poisson model



▶ using ZIP model



## Summary: Model choices

- ▶ It can be concluded (Oliveira et al, 2016):
  - ▶ Zero-inflation is driven by **partial (body) exposure**;
  - ▶ Overdispersion is driven by **densely ionizing radiation**, *i.e.* radiation with high linear energy transfer (LET);

## Summary: Model choices

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  - ▶ Zero-inflation is driven by **partial (body) exposure**;
  - ▶ Overdispersion is driven by **densely ionizing radiation**, *i.e.* radiation with high linear energy transfer (LET);
  - ▶ the absence of partial exposure/ densely ionizing radiation does not guarantee the absence of zero-inflation/overdispersion.
- ▶ Recommended model choices

	exposure		whole body	partial
LET	low		Poisson/NB	ZIP
	high		NB/Neyman A	ZINB

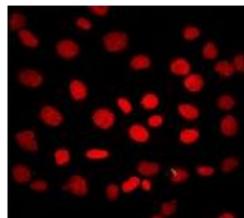
- ▶ Some work has been done to extend dose (uncertainty) estimation routines to NB/ZIP models; further research required (Higuera et al, 2015)

# Strength and limitations of dicentric assay

- ▶ Strength:
  - ▶ very little inter-individual (and inter-laboratory) variation;
  - ▶ dicentric chromosome aberrations can be identified still several weeks after exposure;
  - ▶ internationally accepted 'gold-standard'.
- ▶ Limitations:
  - ▶ needs 3–4 days after irradiation before dicentric chromosomes become visible (need to reach metaphase);
  - ▶ relatively work-intensive and expensive methodology;
  - ▶ not viable for large-scale radiation accidents.
- ▶ Hence, alternative biomarkers have recently been considered.

## $\gamma$ -H2AX data

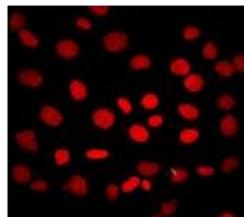
- ▶ Relatively new technology: Protein biomarker
- ▶ Double strand breaks lead to 'phosphorylation' of the H2AX protein, yielding  $\gamma$ -H2AX foci.
- ▶  $\gamma$ -H2AX foci are counted using flow cytometers or immunofluorescence microscopy.
- ▶ Quicker, cheaper, less invasive than dicentric array.





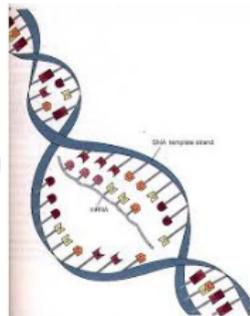
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- ▶ Quicker, cheaper, less invasive than dicentric array.
- ▶ Problems:
  - ▶ only visible up to 24 hours after exposure;
  - ▶ large inter-individual variation, hence requires further calibration steps, or models which can capture this variation.
- ▶ Some ad-hoc dose estimation methods available (Ainsbury et al, 2016).
- ▶ Critical gap remains: Fast ( $< 24h$ ) dose assessment with samples that taken  $> 24h$  after the radiation incident.



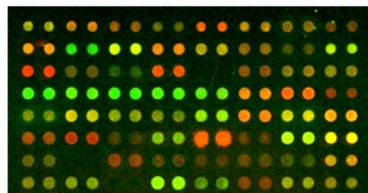
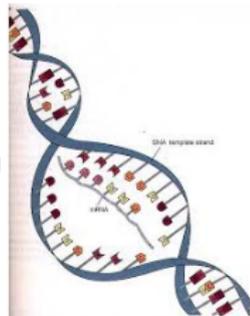
## Microarray-based biomarkers

- ▶ Recently, it has been demonstrated that certain genes respond to ionizing radiation with a change in their gene expression level.
- ▶ Genes are expressed by production of mRNAs from DNA, and protein from mRNAs.
- ▶ mRNA is a linear molecule which carries a copy of the gene to be expressed from the nucleus.



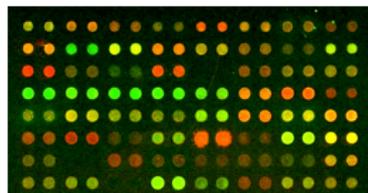
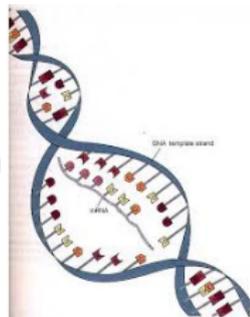
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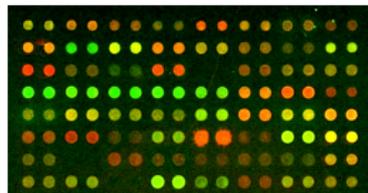
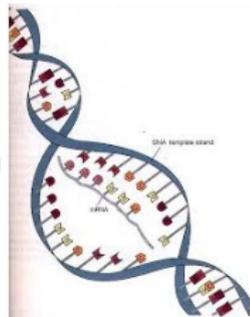
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- ▶ Several radiation-responsive genes are known.
- ▶ Again, substantial inter-individual variation.
- ▶ No reliable statistical methodology for dose estimation yet.



## References

- (1) Merkle W (1983). Statistical methods in regression and calibration analysis of chromosome aberration data. *Radiation and Environmental Biophysics* **21**, 217–233.
- (2) Higuera M, Puig P, Ainsbury EA & Rothkamm K (2015). A new inverse regression model applied to radiation biodosimetry. *Proc's of the Royal Society* **471**, DOI: 10.1098/rspa.2014.0588.
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- (4) Ainsbury EA, Higuera M, Puig P, Einbeck J et al. (2016), Uncertainty of fast biological radiation dose assessment for emergency response scenarios. *International Journal of Radiation Biology*, DOI: 10.1080/09553002.2016.1227106
- (5) Einbeck J & Wilson P (2016). A diagnostic plot for assessing model fit in count data models. In: Dupuys J-F & Josse J (Eds). Proc's of the 31st IWSM, Rennes, France, 4-8/7/2016, 103–108.